

Visualization of Zn²⁺ Ions in Live Zebrafish Using a Luminescent Iridium(III) Chemosensor

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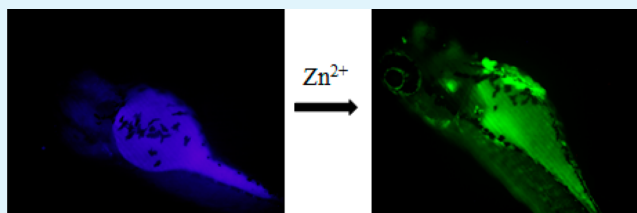
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S Supporting Information

ABSTRACT: A novel luminescent cyclometalated iridium(III) complex-based chemosensor (**1**) bearing a zinc-specific receptor, tris(2-pyridylmethyl)amine, and the 3-phenyl-1*H*-pyrazole ligand has been designed and synthesized. Upon the addition of Zn²⁺ ions to a solution of iridium(III) complex **1**, a pronounced luminescence color change from blue to green can be observed, which may be attributed to the suppression of photoinduced electron transfer upon complexation of complex **1** with Zn²⁺ ions. The interaction of iridium(III) complex **1** with Zn²⁺ ions was investigated by UV–vis absorption titration, emission titration, and ¹H NMR titration. Furthermore, the iridium(III) complex **1** exhibited good selectivity for Zn²⁺ over 13 other common metal ions, including K⁺, Ag⁺, Na⁺, Ni²⁺, Fe³⁺, Hg²⁺, Cd²⁺, Mg²⁺, Ca²⁺, Cu²⁺, Mn²⁺, Co²⁺, and Pb²⁺ ions. The practical application of the iridium(III) complex **1** in visualizing intracellular Zn²⁺ distribution in live zebrafish was also demonstrated.

KEYWORDS: iridium(III) complex, zinc ion, chemosensor, luminescent



1. INTRODUCTION

Zinc ions are involved in important physiological functions in living organisms, such as neurotransmission, gene transcription, and immune function.¹ Zinc deficiency is associated with a number of human diseases, including major depressive disorder, developmental defects, chronic liver disease, chronic renal disease, sickle cell disease, and diabetes.^{2–4} On the other hand, the elevated consumption of zinc has been implicated in ataxia, lethargy, and copper deficiency.^{5,6} Therefore, the accurate detection and imaging of zinc ions is in high demand for the investigation of the regulation of zinc levels and the occurrence of disease states.

A range of detection methods for zinc ion have been developed, including atomic absorption spectrometry⁷ and inductively coupled plasma mass spectrometry.⁸ However, these methods may be time-consuming and/or require the use of sophisticated instrumentation. The notable advantages of luminescence detection have stimulated the development of a variety of fluorescent chemosensors for zinc ions.^{9,10} For example, probes based on zinc-specific receptors conjugated to fluorescent chromophores such as cyanine,¹¹ fluorescein,^{12–23} coumarin,^{24–27} 4-nitrobenzoxadiazole,^{28–30} quinolone,^{31–35} benzoresorufin,³⁶ 1,8-naphthyridine,³⁷ methylazacalix[*a*]pyridine,³⁸ tetraphenylethylene,³⁹ and rhodamine^{40–42} have been reported for Zn²⁺ detection. Some of these fluorescent probes have been used to

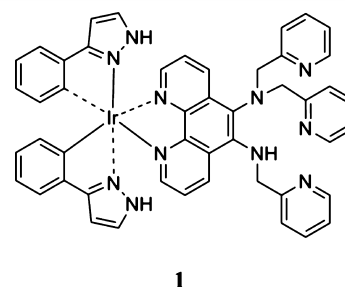


Figure 1. Chemical structure of cyclometalated iridium(III) complex **1**.

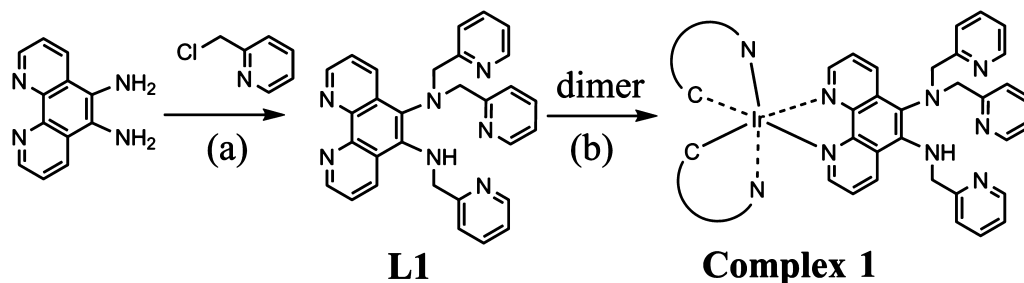
monitor Zn²⁺ in mitochondria of living cells or in the hippocampus slide.^{43,44} However, the use of organic fluorophores in sensing may be limited by their small Stokes shift values and fluorescence lifetimes, as well as autofluorescence arising in biological samples.

In this context, transition-metal complexes with metal-to-ligand charge transfer (MLCT) excited states have arisen as viable alternatives to organic fluorophores and have been widely employed for optoelectronic devices, cellular imaging, and

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Scheme 1. Synthetic Pathway of Iridium(III) Complex 1^a

^aReagents and conditions: (a) picolyl chloride, CH₃CN, K₂CO₃, N₂, reflux, 56.5%; (b) DCM/methanol, reflux.

chemosensing and as structural probes for biomolecules.^{45–84} The long phosphorescence lifetime of luminescent metal complexes in the visible region offers a means to eliminate background fluorescence by the use of time-resolved luminescence spectroscopy. A few luminescent iridium(III) complexes have been developed for Zn²⁺ detection, such as the cyclometalated complexes containing the zinc-specific di-2-picolyamine (DPA) receptor as reported by the groups of Lippard, Nam, You, and Lo.^{85–88} We herein report the application of a novel luminescent cyclometalated iridium(III) complex containing the Zn²⁺ receptor tris(2-pyridylmethyl)amine (TPA) and the 3-phenyl-1H-pyrazole (C[^]N) ligand for Zn²⁺ ion imaging (Figure 1). Furthermore, the iridium(III) complex **1** was demonstrated to function as a selective luminescent probe for visualizing intracellular Zn²⁺ distribution in live zebrafish.

2. EXPERIMENTAL SECTION

2.1. Materials. Reagents, unless specified, were purchased from Sigma-Aldrich (St. Louis, MO) and used as received. Iridium chloride hydrate (IrCl₃·xH₂O) was purchased from Precious Metals Online (Australia).

2.2. Synthesis of Complex 1. The precursor iridium(III) dimer complex Ir₂(C[^]N)₄Cl₂ (where C[^]N = 3-phenyl-1H-pyrazole) was prepared according to modified literature methods.⁸⁹ A suspension of Ir₂(C[^]N)₄Cl₂ (0.2 mmol) and N[^]N ligand **L1** (0.44 mmol) in a mixture of DCM/methanol (1:1, 20 mL) was refluxed overnight under a nitrogen atmosphere. The resulting solution was then allowed to cool to room temperature and filtered to remove unreacted cyclometalated dimer. To the filtrate was added an aqueous solution of ammonium hexafluorophosphate (excess), and the filtrate was reduced in volume by rotary evaporation until precipitation of the crude product occurred. The precipitate was then filtered and washed with several portions of water (2 × 50 mL) followed by diethyl ether (2 × 50 mL). The product was recrystallized by acetonitrile/diethyl ether vapor diffusion to yield the title compound.

Data for **L1**: yield 35%; ¹H NMR (400 MHz, DMSO) δ 9.00 (dd, *J* = 4.2, 1.6 Hz, 2H), 8.76–8.56 (m, 4H), 8.50 (ddd, *J* = 4.8, 1.7, 0.9 Hz, 4H), 7.99 (dd, *J* = 8.2, 1.6 Hz, 4H), 7.80–7.72 (m, 2H), 7.76–7.58 (m, 1H), 7.56–7.37 (m, 2H), 7.30 (ddd, *J* = 7.6, 4.8, 1.1 Hz, 6H), 6.81 (s, 2H), 4.72 (s, 1H); ¹³C NMR (100 MHz, DMSO) δ 158.93, 149.47, 149.03, 148.42, 145.12, 141.35, 136.77, 136.52, 133.21, 130.33, 130.12, 123.29, 122.32, 122.15, 121.85, 120.94, 120.08, 57.90, 54.96; MALDI-TOF-HRMS *m/z* calcd 483.2171, found 484.2260.

Data for complex **1**: yield 35%; ¹H NMR (400 MHz, DMSO) δ 9.15 (dd, *J* = 4.2, 1.4 Hz, 4H), 9.07 (dd, *J* = 4.3, 1.6 Hz, 4H), 8.73–8.66 (m, 4H), 8.63–8.56 (m, 16H), 8.52 (dd, *J* = 8.1, 1.4 Hz, 4H), 8.30 (d, *J* = 5.4 Hz, 3H), 8.26 (s, 3H), 8.16 (s, 1H), 8.06 (d, *J* = 5.4 Hz, 3H), 8.00 (t, *J* = 6.1 Hz, 8H), 7.88–7.84 (m, 3H), 7.80 (dddd, *J* = 8.1, 6.2, 4.0, 2.2 Hz, 12H), 7.72–7.60 (m, 17H), 7.40 (ddd, *J* = 7.6, 4.3, 1.4 Hz, 17H), 7.27 (t, *J* = 7.6 Hz, 3H), 7.03 (td, *J* = 6.2, 2.6 Hz, 3H), 6.95 (t, *J* = 7.6 Hz, 3H), 4.41 (s, 3H), 3.46 (s, 4H); ¹³C NMR (100 MHz, DMSO) δ 164.49, 159.52, 153.31, 151.00, 150.96, 149.10, 147.59, 147.29, 143.53, 136.00, 133.74, 129.17, 127.94, 124.10, 123.38, 123.14, 122.74, 121.49, 119.66,

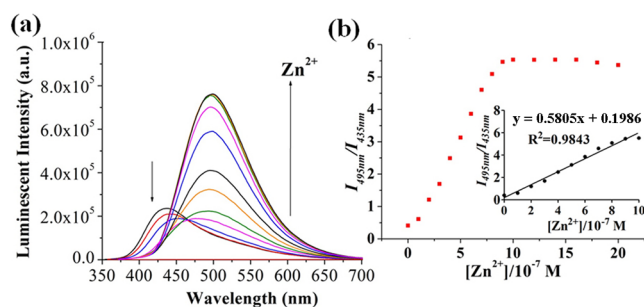


Figure 2. (a) Luminescence spectra of **1** (1 μM) upon addition of various concentrations of Zn²⁺ ions (0–2 μM) in Tris-buffered solution (25 mM, pH 7.04). (b) Relationship between the luminescence intensity and Zn²⁺ concentration. Inset: luminescence response of **1** at I_{495nm}/I_{435nm} vs [Zn²⁺]. λ_{ex} = 320 nm.

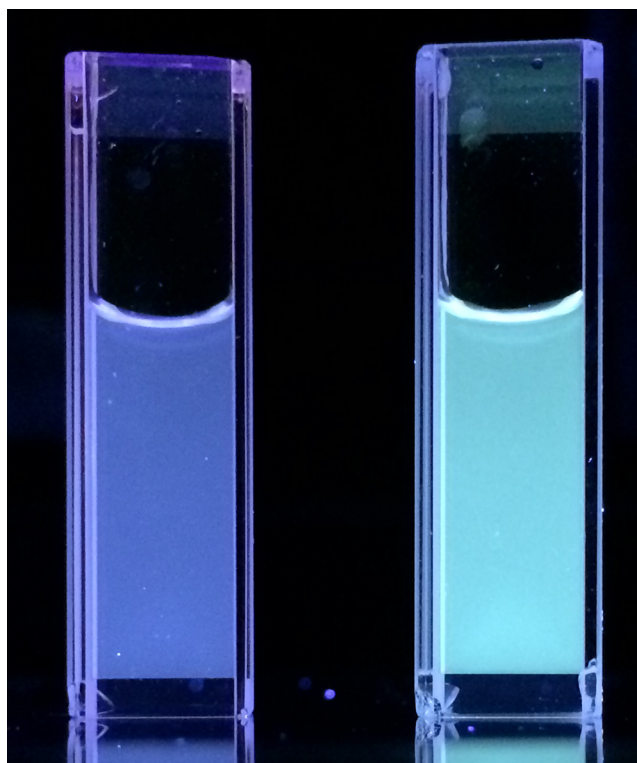


Figure 3. Photograph image of complex **1** (1 μM) in Tris buffer (25 mM, pH 7.04) in the (left) absence or (right) presence of 1 μM Zn²⁺ under UV illumination.

118.64, 117.43, 108.13, 104.95, 98.06, 44.14, 42.52, 36.48, 30.97; MALDI-TOF-HRMS *m/z* calcd 966.3332, found 966.1658.

2.3. Zinc Ion Imaging in Zebrafish. Three day old zebrafish were fed with a solution containing 200 μM Zn^{2+} solution or with a control solution at 28.5 $^{\circ}\text{C}$ for 1.5 h. The zebrafish were washed with PBS solution three times and were incubated with a solution of 40 μM iridium(III) complex **1** at 28.5 $^{\circ}\text{C}$ for 1 h. Alternatively, Zn^{2+} -fed zebrafish were rinsed three times with PBS and then incubated with 50 μM N,N,N',N' -tetrakis(2-pyridylmethyl)ethylenediamine (TPEN) solution, followed by further rinsing three times with PBS and incubation with 40 μM iridium(III) complex **1** at 28.5 $^{\circ}\text{C}$ for 1 h. Confocal imaging of zebrafish was performed using a research inverted microscope system (Olympus IX73).

3. RESULTS AND DISCUSSION

3.1. Design and Synthesis of the Iridium(III) Complex.

The iridium(III) complex **1** was designed to bear a $N^{\wedge}N$ ligand (L1) incorporating the Zn^{2+} receptor TPA,⁹⁰ in addition to two $C^{\wedge}N$ ligands. The coordination of L1 to Zn^{2+} should influence the MLCT state of the complexes, resulting in observable absorption and emission spectral changes of the complex in the presence of Zn^{2+} . Furthermore, the luminescence of the iridium(III) complex is known to be highly sensitive to the

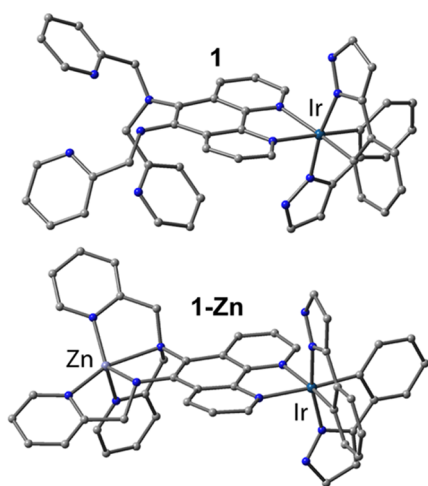


Figure 4. DFT-optimized ground-state structures for **1** and **1**– Zn .

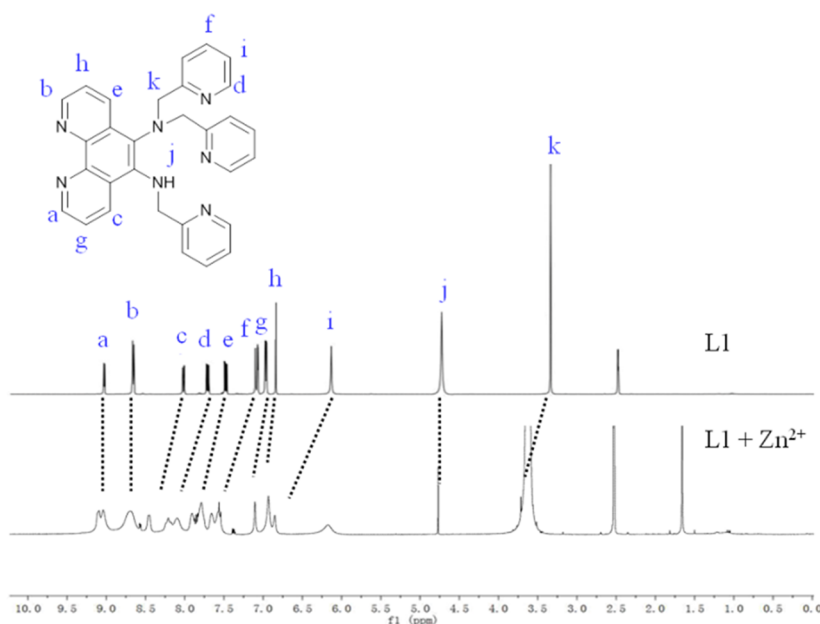


Figure 5. ^1H NMR spectra of L1 (5 mM) in the (upper) absence and (lower) presence of ZnCl_2 (5 mM) in $\text{DMSO}-d_6$ at 298 K.

nature of the ligands. The iridium(III) complex **1** was prepared according to modified literature procedures (Scheme 1).^{89,90}

3.2. Photophysical Response of Complex **1** to Zn^{2+} .

Complex **1** was titrated with different concentrations of Zn^{2+} in Tris buffer (25 mM, pH 7.04). Interestingly, the luminescence of **1** was significantly enhanced in the presence of increasing concentrations of Zn^{2+} ions, which was accompanied by a substantial red shift ($\Delta\lambda = ca. 60$ nm) in the emission maxima of **1** (Figure 2a). This luminescence enhancement is presumably due to the suppression of photoinduced electron transfer upon complexation of complex **1** with Zn^{2+} . A ratiometric ($I_{495\text{nm}}/I_{435\text{nm}}$) luminescence enhancement of *ca.* 6-fold was observed at 2 μM Zn^{2+} ions. Furthermore, a linear relationship was observed between the ratiometric enhancement of complex **1** and the Zn^{2+} concentration ($R^2 = 0.984$) in the range of 0–1 μM Zn^{2+} (Figure 2b). The detection limit at a signal-to-noise ratio of 3 was found to be 36 nM. Furthermore, the presence of Zn^{2+} could be readily observed by the naked eye under UV illumination (Figure 3). In the UV–vis spectrum, the addition of Zn^{2+} to complex **1** led to an increase in absorption at 261 and 298 nm and a decrease in absorption at 281 nm (Figure S1, Supporting Information). Additionally, Job's plot analysis of the luminescence data revealed a maximum in emission enhancement at a 0.5 mole fraction of **1**, indicating a 1:1 stoichiometry between Zn^{2+} ions and **1** (Figure S2, Supporting Information).

3.3. Theoretical Studies. Ir(III) complexes are known to possess rich electronic excited states. Time-dependent density functional theory (TD-DFT) calculations have been performed on complex **1** and a hypothetical Zn^{2+} -coordinated complex **1**– Zn to investigate the nature of the excited states. The optimized ground-state structures for **1** and **1**– Zn are depicted in Figure 4; their simulation spectra are depicted in Figure S3 (Supporting Information). Because the nature of the transitions in the spectral region of 300–600 nm are quite complex for both complexes, only the major transitions with oscillator strength >0.04 are presented in Figure S3. For complex **1**, the calculated lowest energy absorption bands at around 425 nm are attributed to a mixing of $n(\text{amine}) \rightarrow \pi^*(\text{phen})$ and $d_{\pi}(\text{Ir}) \rightarrow \pi^*(\text{phen})$ charge transfer transitions (transitions I and II in Figure S4, Supporting Information).

For hypothetical complex **1**–Zn, the calculated lowest energy absorption band at around 475 nm is attributed to the $d_{\pi}(\text{Ir}) \rightarrow \pi^*(\text{phen})$ charge transfer transition (transition I in Figure S5, Supporting Information). The calculated red shift upon Zn^{2+} coordination to **1** is consistent with the experimental finding that a red shift in emission energy is induced upon Zn^{2+} binding to **1**.

3.4. ^1H NMR Titration. To investigate the complexation mode of Zn^{2+} ions, ^1H NMR titration of **L1** with Zn^{2+} in $\text{DMSO}-d_6$ was performed. The results showed several significant spectral changes in the ^1H NMR spectrum of **L1** upon the addition of Zn^{2+} ions (c, from 7.98 to 8.41 ppm; d, from 7.72 to 8.19 ppm; e, from 7.46 to 7.82 ppm; f, from 7.13 to 7.64 ppm; g, from 6.94 to 7.17 ppm; h, from 6.63 to 6.90 ppm; i, from 6.17 to 6.65 ppm; k, from 3.31 to 3.72 ppm) (Figure 5). The spectral changes suggest that Zn^{2+} ions putatively bind to **L1** via coordination to the nitrogen atoms of the pyridine groups.

3.5. Selectivity of Iridium(III) Complex **1 for Zn^{2+} .** We next evaluated the selectivity of the iridium(III) complex **1**

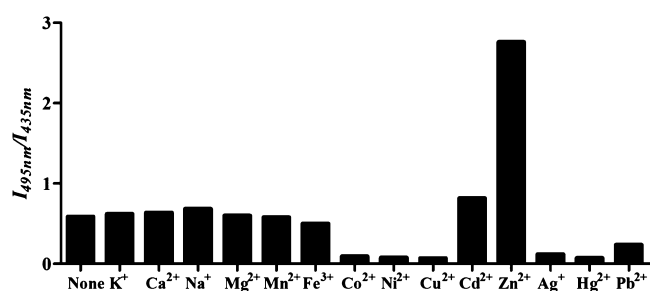


Figure 6. Selectivity of complex **1** for Zn^{2+} . The concentration of Zn^{2+} ions was $1.0 \mu\text{M}$, the concentrations of Na^+ and K^+ were $1000 \mu\text{M}$, and the concentrations of the other metal ions were $20.0 \mu\text{M}$.

for Zn^{2+} by investigating its luminescence response to 13 other common metal ions, including Na^+ , K^+ , Ag^+ , Ni^{2+} , Fe^{3+} , Hg^{2+} , Cd^{2+} , Mg^{2+} , Ca^{2+} , Cu^{2+} , Mn^{2+} , Co^{2+} , and Pb^{2+} ions. The results showed that only Zn^{2+} ions could significantly enhance the ratiometric luminescence output of complex **1** (Figure 6). By comparison, only minor changes in $I_{495\text{nm}}/I_{435\text{nm}}$ were observed upon the addition of a 20-fold excess of the Ag^+ , Ni^{2+} , Fe^{3+} , Hg^{2+} , Cd^{2+} , Mg^{2+} , Ca^{2+} , Cu^{2+} , Mn^{2+} , Co^{2+} , and Pb^{2+} ions. Moreover, complex **1** displayed significant selectivity for Zn^{2+} ions over a 1000-fold excess of Na^+ and K^+ ions. These results indicate that the iridium(III) complex **1** displays significant selectivity for Zn^{2+} ions over the other metal ions, which originates presumably from the specific interaction of the TPA moiety of complex **1** with Zn^{2+} ions. Additionally, we have investigated the impact of the coaddition of Cu^{2+} ions on the performance of the assay for Zn^{2+} detection. The results showed that addition of 0.2 or $0.4 \mu\text{M}$ Cu^{2+} into a $1 \mu\text{M}$ Zn^{2+} solution would result in a 24% or 33% decrease in the luminescence of complex **1**, respectively (Figure S6, Supporting Information).

3.6. Zn^{2+} Imaging in Zebrafish. The lifetime of complex **1** was determined to be $4.38 \mu\text{s}$ ($\lambda = 495 \text{ nm}$), which could make it suitable for Zn^{2+} imaging in biological tissues. Taking advantage of the ratiometric behavior and high sensitivity of complex **1** toward Zn^{2+} ions, the practical application of complex **1** as a luminescent probe to monitor Zn^{2+} distribution in live zebrafish was investigated. Ratiometric imaging revealed a bright green fluorescence in the abdomen of 3 day old zebrafish fed with Zn^{2+} , indicating an accumulation of Zn^{2+} ions in the abdomen, while non- Zn^{2+} -treated zebrafish appeared blue (Figure 7). Comparison of the luminescence images obtained for zebrafish fed with different concentrations of zinc ions (0, 20 nM, 200 nM, $2 \mu\text{M}$, $20 \mu\text{M}$, and $200 \mu\text{M}$) suggested that down to 200 nM Zn^{2+} ions

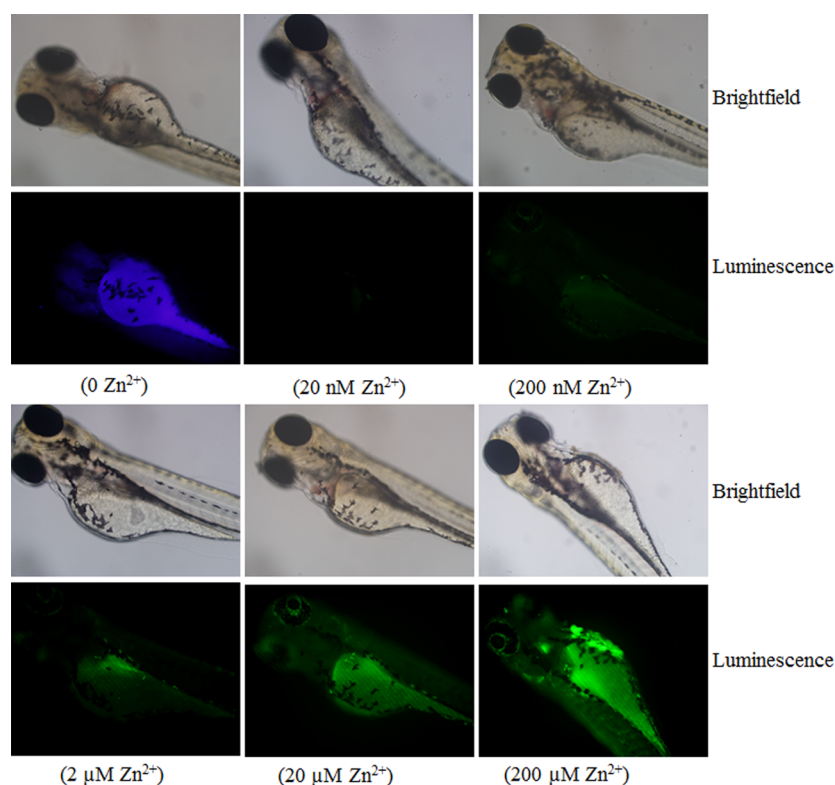


Figure 7. Zn^{2+} imaging in 3 day old zebrafish. Bright-field (upper) and luminescence (lower) images of zebrafish fed with different concentrations of Zn^{2+} (0, 20 nM, 200 nM, $2 \mu\text{M}$, $20 \mu\text{M}$, and $200 \mu\text{M}$) for 1.5 h followed by incubation with complex **1** ($40 \mu\text{M}$) for 1 h.

could be detected in live zebrafish (Figure 7). A time-course experiment revealed that the green fluorescence intensity of treated zebrafish increased with the Zn^{2+} incubation time (0, 30, 60, and 90 min) (Figure S7, Supporting Information). Additionally, the green fluorescence in Zn^{2+} -treated zebrafish was reduced to an almost undetectable level if the chelator TPEN (50 μM) for 1.5 h and incubation with complex 1 (40 μM) for 1 h.

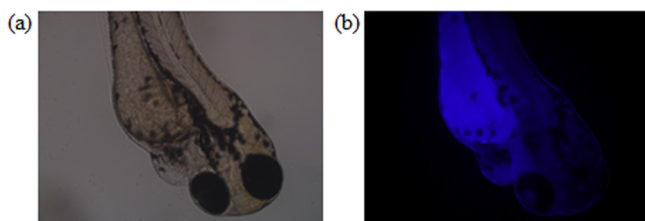


Figure 8. (a) Bright-field and (b) luminescence images of a zebrafish incubated with Zn^{2+} (200 μM) for 1.5 h, followed by treatment with TPEN (50 μM) for 1.5 h and incubation with complex 1 (40 μM) for 1 h.

observed in Figure 7 arose from the complexation of complex 1 with Zn^{2+} . These results indicate that complex 1 can be used as a luminescent imaging agent for Zn^{2+} ions in live zebrafish.

4. CONCLUSION

In summary, we have synthesized a novel luminescent cyclometalated iridium(III) complex containing the zinc-binding TPA group. Complex 1 displayed pronounced luminescence changes upon the addition of Zn^{2+} in aqueous solution and showed good sensitivity and selectivity for Zn^{2+} over 13 other metal ions, including K^+ , Ag^+ , Na^+ , Ni^{2+} , Fe^{3+} , Hg^{2+} , Cd^{2+} , Mg^{2+} , Ca^{2+} , Cu^{2+} , Mn^{2+} , Co^{2+} , and Pb^{2+} ions. Compared to other organic zinc ion chemosensors, which typically require an organic solvent for detection, our developed iridium(III) complex-based Zn^{2+} chemosensor could be used in aqueous buffer and exhibits a higher selectivity against Cd^{2+} ions than that of some other reported Zn^{2+} ion chemosensors. A comparison of recently reported chemosensors for Zn^{2+} ion is presented in Table 1. Furthermore, the

Table 1. Comparison of Chemosensors for Zn^{2+} Ion Assays Recently Reported in the Literature

signal output	detection condition	selectivity against Cd^{2+}	real sample application	ref
ratiometric and colorimetric	ACN/ H_2O (80:20)	good	not reported	90
fluorescent	HEPES buffer	not good	HeLa cells	91
fluorescent	ACN/ H_2O (18:7)	good	not reported	92
fluorescent	ACN	good	not reported	93
fluorescent	DMF/ H_2O (1:1)	not good	not reported	94
fluorescent	EtOH/ H_2O (9:1)	not good	HepG-2 cells	95
fluorescent	EtOH	good	MCF-7 cells	96
luminescent and ratiometric	Tris buffer	good	zebrafish	this study

practical application of complex 1 for imaging Zn^{2+} ions in live zebrafish was successfully demonstrated. We envision that the iridium(III) complex could be further developed as a useful probe for monitoring zinc ion distribution in biological samples.

■ ASSOCIATED CONTENT

Supporting Information

General experimental procedures, photophysical measurements, DFT calculations, UV-vis absorption spectra, and Job's plot

analysis. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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